

Enantioselective Recognition of Amino Acids by Axially-Chiral π -Electron-Deficient Receptors

Masumi Asakawa,[†] Christopher L. Brown,[†] Dario Pasini,[†] J. Fraser Stoddart,^{*,†} and Paul G. Wyatt[‡]

School of Chemistry, University of Birmingham, Edgbaston, Birmingham B15 2TT, U.K., and Glaxo Wellcome Medicines Research Centre, Gunnels Wood Road, Stevenage, Hertfordshire SG1 2NY, U.K.

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The development of chiral hosts capable of exhibiting enantiomeric differentiation toward racemic guests has become an important and rapidly growing field of chemistry.¹ Recent work has led to the design and syntheses of chiral hosts for alkaloids,² amino acids,³ peptides,⁴ and other neutral guests.⁵ Meanwhile, cyclobis(paraquat-*p*-phenylene) (**1**⁴⁺), which has been self-assembled⁶ around numerous cyclic and acyclic templates to produce catenanes and rotaxanes,⁷ has been shown^{8,9} —as its tetrakis(hexafluorophosphate) salt **1**·4PF₆[−] to complex (Figure 1) with π -electron-rich substrates like 1,4-bis(2-(2-hydroxyethoxy)ethoxy)benzene (BHEEB) strongly in organic solvents.

Furthermore, as its tetrachloride salt **1**·4Cl in H₂O, **1**⁴⁺ forms strong 1:1 complexes with biologically-important compounds like π -electron-rich aromatic amino acids,¹⁰ a series of catechol-containing neurotransmitters,¹¹ and phenyl glycopyranosides.¹² One of its main features as

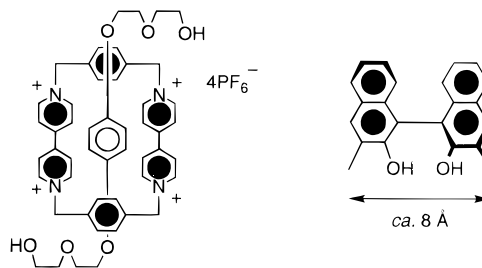


Figure 1. 1:1 complex between **1**·4PF₆ and BHEEB (on the left) and the rigid, axially-chiral binaphthol spacer (see text for details).

a host¹³ is its rigid rectangular cavity wherein the distance¹³ between the π -electron-deficient bipyridinium units (7 Å) is ideal for the inclusion of a π -electron-rich aromatic ring. In order to provide ourselves with optically-active derivatives of **1**⁴⁺ that retain its receptor characteristics, we decided to investigate introducing axially-chiral 3,3'-linked-2,2'-dihydroxy-1,1'-binaphthyl spacers sequentially, in place of the *p*-phenylene rings, into the tetracationic cyclophane. Not only should it hold the two bipyridinium units *ca.* 8 Å apart, but it is also among the most versatile and popular chiral building block with C₂ symmetry used¹⁴ in the resolution of enantiomers and in asymmetric syntheses and catalyses. In this paper, we (1) describe the template-directed synthesis (Scheme 1) of the new axially-chiral tetracationic cyclophanes (*R*)-**4**·4PF₆ and (*RR*)-**6**·4PF₆ and (2) report on their abilities to differentiate between the D- and L-enantiomers of the aromatic amino acids phenylalanine, tyrosine, and tryptophan, in H₂O and their N-acetylated derivatives in organic solvents.

The synthesis of the optically-active dibromide (*R*)-**2** was carried out as described previously.¹⁵ The tetracationic cyclophane (*R*)-**4**·4PF₆ was self-assembled by stirring equimolar amounts of (*R*)-**2** and **3**·2PF₆⁹ in the presence of an excess of BHEEB for 14 days at room temperature. The highly colored reaction mixture was purified⁹ by liquid–liquid extraction (CHCl₃/H₂O), followed by column chromatography (SiO₂; MeOH–2 M NH₄Cl–MeNO₂, 7:2:1) and counterion exchange (NH₄PF₆, H₂O). The crystalline product was recrystallized from MeCN/*i*-Pr₂O to give the optically-active-[α]_D²⁵ = +86° (*c* = 0.009, Me₂CO)–C₂ symmetrical¹⁶ receptor (*R*)-**4**·4PF₆ in 27% yield. Employing similar isolation and purification techniques, the optically-active-[α]_D²⁵ = +103° (*c* = 0.006, MeCN)–D₂ symmetrical¹⁶ receptor (*RR*)-**6**·4PF₆ was self-assembled by mixing equimolar amounts of (*R*)-**2** and (*R*)-**5**·2PF₆¹⁷ in the presence of an excess of the

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(16) The symmetry elements present in (*R*)-**4**·4PF₆ and (*RR*)-**6**·4PF₆ mean that in both cases the faces of the receptors are homotopic. Thus, irrespective of which face binds the substrate, only one 1:1 complex can be formed by (*R*)-**4**·4PF₆ and (*RR*)-**6**·4PF₆.

[†] University of Birmingham, U.K.

[‡] Glaxo Wellcome Medicines Research Centre, U.K.

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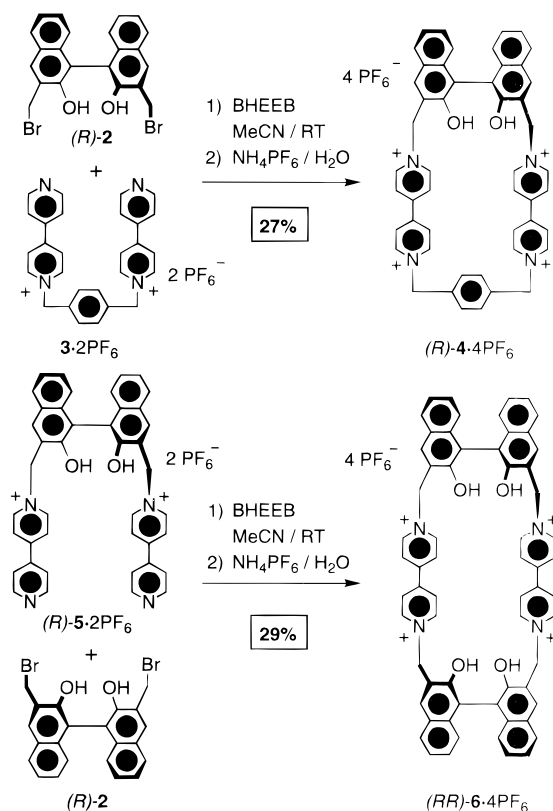
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Table 1. Binding Constants (K_a) and Free Energies of Complexation ($-\Delta G^\circ$) for the 1:1 Complexes between Cyclophanes (R)-4**-X (X = PF₆ or Cl) and (RR)-**6**-4PF₆ and π -Electron-Rich Amino Acids^a**

entry	substrate	solvent	K_2 (M ⁻¹)		$K_a(L)/K_a(D)$		$-\Delta G^\circ$ (kcal/mol)		$\Delta\Delta G^\circ$ (kcal/mol) ^b	
			(R)- 4	(RR)- 6	(R)- 4	(RR)- 6	(R)- 4	(RR)- 6	(R)- 4	(RR)- 6
1	L-Try	H ₂ O ^c	2470	ND ^d			4.63			
2	D-Try	H ₂ O ^c	5860	ND ^d	0.42		5.14		-0.51	
3	L-Try OMe·HCl	H ₂ O ^c	753	ND ^d			3.92			
4	D-Try OMe·HCl	H ₂ O ^c	803	ND ^d	0.94		3.96		-0.04	
5	<i>N</i> -Ac-L-Try	A ^e	20700	4280			5.89	4.95		
6	<i>N</i> -Ac-D-Try	A ^e	2670	1080	7.75	3.96	4.67	4.14	1.22	0.81
7	<i>N</i> -Ac-L-Tyr	B ^e	10060	2340			5.45	4.60		
8	<i>N</i> -Ac-D-Tyr	B ^e	2125	1047	4.73	2.23	4.53	4.12	0.92	0.48
9	<i>N</i> -Ac-L-Phe	A ^e	1220	219			4.21	3.19		
10	<i>N</i> -Ac-D-Phe	A ^e	2260	137	0.54	1.60	4.57	2.91	-0.36	0.28

^a All binding constants were determined by UV/vis titration at 25 °C. ^b $\Delta\Delta G^\circ = \Delta G^\circ(L) - \Delta G^\circ(D)$. ^c In H₂O as a solvent, (R)-**4**-4Cl was used. ^d Not determined. ^e Solvent mixture A: MeCN 90% DMF 10%; solvent mixture B: MeCN 90% DMSO 10%.

Scheme 1

template (BHEEB) in 29% yield. Both receptors gave good elemental analyses and were characterized by LSIMS and ¹H-NMR and ¹³C-NMR spectroscopies.

Although inspection of molecular models show that (R)-**4**⁴⁺ and (RR)-**6**⁴⁺ have slightly enlarged and distorted cavities compared with **1**⁴⁺, both chiral tetracationic cyclophanes form strong 1:1 complexes with BHEEB: for (R)-**4**·4PF₆ and (RR)-**6**·4PF₆ in MeCN at 25 °C, the K_a values are 810 and 2640 M⁻¹, respectively, as determined by UV/vis titration. Binding constants and derived free energies of complexation for the 1:1 complexes formed between either (R)-**4**-4X (X = PF₆ or Cl)¹⁸ or (RR)-**6**·4PF₆ and the L- and D-enantiomers of phenylalanine (Phe), tyrosine (Tyr), and tryptophane (Try)—as free, methyl esterified, or *N*-acetylated—in H₂O and organic solvents are listed in Table 1. Comparison of the K_a values

between (R)-**4**·4Cl and L-Try and D-Try in H₂O (Table 1, entries 1 and 2) indicate a modest amount of enantioselectivity in favor of D-Try that was reduced drastically when the corresponding L- and D-methyl ester hydrochloride salts were investigated (Table 1, entries 3 and 4), suggesting that the carboxylic acid function in these amino acids might be playing a role in binding and enantioselection. When the amino groups on L-Try and D-Try are *N*-acetylated, the enantioselectivity in MeCN/DMF (90:10) is even larger but favors the L- over the D-enantiomer (Table 1, entries 5 and 6). In both cyclophanes, the enantioselectivities decrease on going from *N*-acetyltryptophan to *N*-acetyltyrosine¹⁹ (Table 1, entries 7 and 8) and to the less π -electron-rich *N*-acetyl phenylalanine (Table 1, entries 9 and 10). All these data suggest to us a model, where the more π -electron rich is the guest (primary mode of binding), the more the secondary stereoelectronic interactions of the functional groups of the guest with the bulky optically-active binaphthol spacer(s) of the cyclophane(s) are effective²⁰ in recognition terms. Indeed, on comparing the 1:1 complexes between *N*-acetyl L- and D-tryptophan and (R)-**4**⁴⁺, molecular mechanics Monte Carlo conformational searching²¹ using Macromodel 5.0 (1000 structures, OPLS force field, and the GB/SA solvation model for water) predicts that the L-enantiomer complex will be more stable by approximately 1.5 kcal/mol compared with the D-enantiomer complex. A fully systematic molecular modeling study aimed at further interpreting the observations recorded in Table 1 is underway currently.

We believe that the axially-chiral π -electron-deficient receptors (R)-**4**⁴⁺ and (RR)-**6**⁴⁺ can be adapted (1) to effect the resolutions of racemic substrates containing π -electron-rich aromatic rings and also (2) to act as asymmetric catalysts of appropriate reactions performed on suitably bound and oriented substrates.

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Supporting Information Available: Experimental section and spectroscopic data for compounds (R)-**4**·4PF₆, (R)-**5**·2PF₆, and (RR)-**6**·4PF₆ and curves obtained from selected UV/vis titrations (5 pages).

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(17) Compound (R)-**5**·2PF₆ was prepared in a manner similar to that for **3**·2PF₆. See ref 9.

(18) Compound (R)-**4**·4Cl was obtained by adding a saturated solution of *n*-Bu₄NCl in Me₂CO to a solution of (R)-**4**·4PF₆ in Me₂CO until no further precipitation occurred.

(19) *N*-Acetyl-D-tyrosine was prepared according to: Du Vigneaud, V.; Meyer, C. E. *J. Biol. Chem.* **1933**, *97*, 295–308.

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